Gas chromatographic-mass spectrometric confirmation of 19-nortestosterone in the urine of untreated boars — effect of the administration of Laurabolin®

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SUMMARY The presence of 17β -19-nortestosterone (nandrolone, NT, 17β -19-NT) and its epimer 17α -19-nortestosterone (epiNT, 17α -19-NT) was investigated in the urine of six untreated boars, obtained from experimental farms. The presence of 17β -19-nortestosterone was screened by RIA and HPTLC and confirmed by GC-MS analysis. Additionally, the two epimers (NT and epiNT) were investigated in the urine of a boar (two-year-old miniature male pig weighing 50 kg) before and after injection of 100 mg Laurabolin® (nortestosterone laurate, Intervet N.V., Belgium).

The isolation of the steroids was based on sample clean-up with solid phase extraction and subsequent high-performance liquid chromatography. Both gas chromatographic retention data and mass spectrometric data (selected ion monitoring and full spectrum) were used for detection and identification.

The presence of 17β -19-nortestosterone in the urine of the boars that were not injected proves the endogenous production of the steroid. The absence of the 17α -epimer in the urine of the injected boar suggests that 17α -19-nortestosterone is not a major metabolite of 17β -19-nortestosterone.

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INTRODUCTION

The illegal use of anabolic steroids in livestock breeding has taken on enormous proportions in the last few decades. Measures are taken by the health authorities to protect the consumer from possible harmful effects due to the consumption of contaminated meat or meat products. Detection of the illegal use of anabolics is difficult as a result of the extensive metabolism of steroid hormones and the possibility of interference with endogenous steroids, which are species dependent. Nandrolone (17 β -19-nortestosterone, NT, 17 β -19-NT) is a frequently used anabolic steroid in cattle fattening. However, 19-nortestosterone has also been detected as a natural steroid in stallion urine (8, 9), it has been identified in the follicular fluid of the mare (4, 13) and its secretion by the testis of the stallion (1, 5) and the boar (11) has been evidenced. In human follicular fluid 19-nortestosterone has also been identified (3, 4). Only some of the above references specify the 17 β -epimeric form of 19-nortestosterone.

Nandrolone has also been detected and quantified in edible parts of non-castrated male pigs, in particular in the liver and muscle tissue (15). Besides boar taint, which is due to the presence of androstenone (5α -androst-16-en-3-one), the occurrence of 17β -19-nortestosterone as a natural constituent of boar meat produces a second 'steroid problem' in pork trade in Europe. Moreover, the tendency to reduce boar castration in the EEC will increase these problems further. Nandrolone will become a more likely natural constituent of pork and meat products derived from pork. Recently, 17β -19-NT was detected in ravioli imported from Italy into Germany. Since the pork used in these products could be originating from non-castrated boars, 17β -19-NT should be considered to be natural in ravioli. This is extremely important for food inspection services.

During regulatory control in the slaughterhouse, it was frequently observed that boar urine shows a RIA-positive result on 19-nortestosterone. The presence of 17β -19-NT in these urine samples was already presumed by one of the authors and later confirmed by some Belgian laboratories involved in the control for the Institute of Veterinary Inspection (10, Van Peteghem C, De Brabander HF, Smits F, Pottie G. Unpublished results). In order to demonstrate that residues could be due to endogenous production, urine samples of clean boars (n = 6) from experimental farms were analysed. Additionally, the presence of 17β -19-nortestosterone and its epimer 17α -19-nortestosterone (epiNT, 17α -19-NT) was traced in the urine of a two-year-old miniature male pig after treatment with Laurabolin®

MATERIALS AND METHODS

(nortestosterone laurate).

Procedure

In the present study, gas chromatography-mass spectrometry (GC-MS) was used for the confirmation of 17β -19-NT after the preliminary radioimmunoassay (RIA) (6) and high-performance thin-layer chromatography (HPTLC) (14) screening tests.

Animals

Urine samples were obtained from six untreated boars (Belgian Landrace), obtained from experimental farms, having an age between two and three years and weighing between 150 and 200 kg. A two-year-old miniature boar (75% Göttinger, 25% Vietnamese) of 50 kg was treated with Laurabolin[®] (100 mg nortestosterone laurate, Intervet N.V. Belgium). Urine samples were collected before and at regular intervals after Laurabolin[®] treatment.

Instrumentation

Solid-phase extraction for sample preparation was carried out using octadecyl (C₁₈) and amino (NH₂) disposable columns (J. T. Baker, Phillipsburg, New Jersey, USA).

High-performance liquid chromatography (HPLC) was performed using a Waters pump (model 6000 A, Waters Associates, Milford, Massachusetts, USA) equipped with an automatic injector (WISP 710 B, Waters Associates, Milford, Massachusetts, USA) and a variable wavelength detector (model SP 8400, Spectra Physics, Santa Clara, California, USA). The HPLC analytical column used was a LiChrospher® 100 RP — 18 (5 μ m) column (Merck, Darmstadt, FRG) and was protected by a guard column (75 mm x 2.1 mm, cat. no. 28603, Chrompack, Middelburg, The Netherlands).

Gas chromatographic-mass spectrometric analyses were carried out on a HP-5890 gas chromatograph equipped with a 25 m HP Ultra 2 (Cross-Linked 5% Phenyl Methyl Silicone — 25 m x 0.2 mm I.D. x 0.33 µm film thickness) fused silica capillary column and linked to a HP-5970 mass-selective detector (Hewlett-Packard, Palo Alto, California, USA).

Sample preparation

Steroid extraction was based on the procedure of Schmidt et al. (12).

Free and conjugated steroids, contained in 10 mL of urine, were extracted on a C_{18} -column. The conjugated steroids were hydrolysed by means of Helix pomatia digestive juice (Boehringer, Mannheim, FRG) at 37°C for 16 hours. The steroids were subsequently extracted on a C_{18} -column. Additional purification was done by placing an amino column (NH₂) in series with the C_{18} -column.

The extract was finally fractionated by means of HPLC (mobile phase CH₃OH/H₂O: 65/35 at a flow-rate of 1 mL/min). The fraction containing the two epimers (5—7.5 min) was collected and evaporated to dryness under nitrogen.

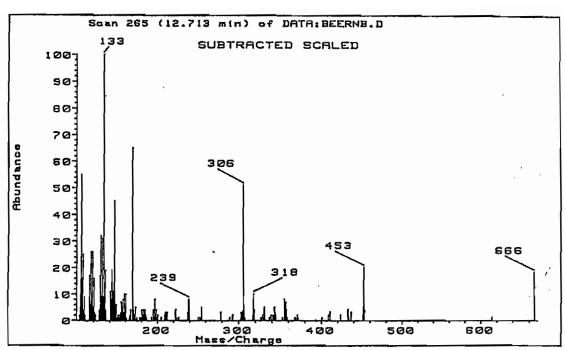
GC-MS analysis

After derivatisation with heptafluorobutyric acid anhydride (HFBA, Macherey-Nagel, Düren, FRG), an aliquot was injected into the GC-MS instrument. The injection temperature and the transfer line temperature were 280°C. The oven temperature was programmed from 200 to 270°C at 5°C/min and was kept constant at 270°C for 14 minutes. The carrier gas was helium at a column head pressure of 80 kPa. Samples were injected in the split mode. The mass spectrometer was used in the electron impact mode.

RESULTS AND DISCUSSION

During preliminary experiments, the presence of 17β -19-NT was demonstrated in the urine of the clean boars (n = 6) by RIA and HPTLC. These results were confirmed by GC-MS in the selected ion monitoring (SIM)-mode. Ion chromatograms of the molecular ion (m/z 666) and two fragment ions (m/z 453 and m/z 306) were recorded. For a positive identification, the three ions must appear simultaneously at the right retention time, expressed in methylene unit values, as derived from the retention data of the pure reference compounds. The methylene unit values of 17α -19-NT-diHFB and 17β -19-NT-diHFB are 23.62 and 24.38, respectively. These results give evidence of the endogenous production of 17β -19-NT in boars. Due to the apparently high concentration at which 17β -19-NT occurs in boar urine, the mass-selective detector could be used in the SCAN-mode, so that a full mass spectrum and thus more scientific evidence can be obtained. As can be seen from Figure 1, the spectrum of the alleged 17β -19-NT-diHFB (top). is identical to that of the reference compound 17β -19-NT-diHFB (bottom). No 17α-19-NT could be detected in the samples taken before Laurabolin® administration, nor could any be detected in the urine samples of the miniature boar taken 92 hours, one week, two weeks and three weeks after the injection of Laurabolin[®]. The method was validated by spiking urine samples with 17α -19-NT in a concentration of 5 ppb. It proves that, in contrast to what is observed in cattle (2, 7, 16), C-17 epimerisation is not a major pathway of metabolism of 17β -19-NT in boars.

In the future, experiments will be set up to investigate 19-nortestosterone metabolites in castrated and female pigs and to expand the results using a larger population of experimental animals.



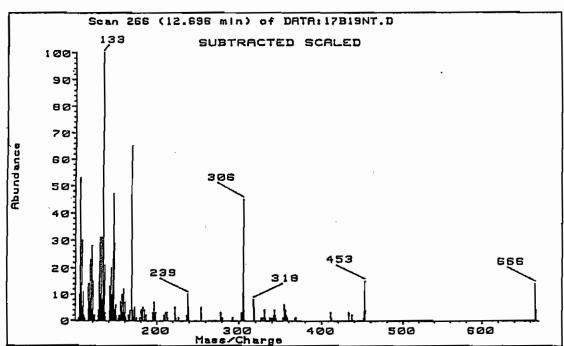


Figure 1. Mass spectrum of 17β -19-NT-diHFB in the untreated boar (top) and of the reference 17β -19-NT-diHFB (bottom).

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REFERENCES

1. Benoît E, Garnier F, Courtot D, and Delatour P. Radioimmunoassay of 19-nortestosterone. Evidence of its secretion by the testis of the stallion. Ann Rech Vét 1985; 16: 379-83.

- 2. Benoît E, Guyot JL, Courtot D, and Delatour P. Gas chromatographic/mass spectrometric analysis of 19-nortestosterone urinary metabolites in cattle. Ann Rech Vét 1989; 20: 485-91.
- 3. Dehennin L, Jondet M, and Scholler R. Androgen and 19-norsteroid profiles in human preovulatory follicles from stimulated cycles: an isotope dilution-mass spectrometric study. J Steroid Biochem 1987; 26: 399-405.
- 4. Dehennin L, Silberzahn P, Reiffsteck A et Zwain I. Présence de 19-norandrostènedione et de 19-nortestostérone dans les fluides folliculaires humain et équin. Pathol Biol 1984; 32: 828-9.
- 5. Dintinger T, Gaillard JL, Zwain I, Bouhamidi R, and Silberzahn P. Synthesis and aromatisation of 19-norandrogens in the stallion testis. J Steroid Biochem 1989; 32: 537-44.
- Evrard P, Gaspar P, and Maghuin-Rogister G. A specific radioimmunoassay for the detection of 19-nortestosterone residues in urine and plasma of cattle. J Immunoassay 1986; 7: 353-63.
- 7. Haasnoot W, Schilt R, Hamers ARM, Huf FA, Farjam A, Frei RW, and Brinkman UATR. Determination of β -19-nortestosterone and its metabolite α -19-nortestosterone in biological samples at the sub parts per billion level by high-performance liquid chromatography with online immunoaffinity sample pretreatment. J Chromatogr 1989; 489: 157-71.
- 8. Houghton E, Copsey J, Dumasia MC, Haywood PE, Moss MS, and Teale P. The identification of C-18 neutral steroids in normal stallion urine. Biomed Mass Spectrom 1984: 11: 96-9.
- 9. Houghton E, Dumasia MC, Moss MS, and Teale P. 19-Nortestosterone: its identification as an endogenous steroid in the colt and the resultant problems of control. Proceedings of the 37th and 38th Meetings of the AORC, Toronto & Denver, 1983 & 1984; 61-71.
- Maghuin-Rogister G, Bosseloire A, Gaspar P, Dasnois C et Pelzer G. Identification de la 19nortestosterone (nandrolone) dans l'urine de verrats non castrés. Ann Méd Vét 1988; 132: 437-40.
- 11. Ruokonen A and Vihko R. Steroid metabolism in testis tissue: concentrations of unconjugated and sulfated neutral steroids in boar testis. J Steroid Biochem 1974; 5: 33-8.
- Schmidt NA, Borburgh HJ, Penders TJ, and Weykamp CW. Steroid profiling An update. Clin Chem 1985; 31: 637-9.
- 13. Silberzahn P, Dehennin L, Zwain I, and Reiffsteck A. Gas chromatography-mass spectrometry of androgens in equine ovarian follicles at ultrastructurally defined stages of development. Identification of 19-nortestosterone in follicular fluid. Endocrinology 1985; 117: 2176-81.
- 14. Smets F. Method of analysis for the determination of residues of anabolics in urine by HPLC-HPTLC. Document of the Benelux Economic Union SP/LAB/h (88) 33.
- Ginkel LA van, Stephany RW, Zoontjes PW, Rossum HJ van, Blitterswijk H van, and Zuijdendorp J. The presence of nortestosterone in edible parts of non-castrated male pigs. Tijdschr Diergeneeskd 1989; 114: 311-4.
- Ginkel LA van, Stephany RW, Rossum HJ van, Blitterswijk H van, Zoontjes PW, Hooijschuur RCM, and Zuydendorp J. Effective monitoring of residues of nortestosterone and its major metabolite in bovine urine and bile. J Chromatogr 1989; 489: 95-104.